# THE EFFECT OF CHRONIC CARBON TETRACHLORIDE INHALATION ON THE SECRETION OF LIPID BY RAT LIVER\*

# LEWIS KANICS and DAVID RUBINSTEIN

Department of Biochemistry, McGill University, Montreal, Canada

(Received 12 January 1968; accepted 23 February 1968)

Abstract—The effect on hepatic metabolism of chronic daily 8-hr exposures of rats to 68 or 680 ppm CCl<sub>4</sub> vapor was studied. After 6 days of exposure there is an elevation of serum glutamic oxaloacetic transaminase and hepatic triglyceride concentration and a decrease in hepatic glycogen levels. Intravenous administration of palmitate-<sup>14</sup>C at the end of the fifth daily exposure period to 68 ppm CCl<sub>4</sub> revealed a decrease in the rate of release of labeled lipid from the liver when compared to that in control animals. This was reflected in a decrease in the amount of labeled triglycerides in the serum of the intoxicated animals. After exposure to 680 ppm for 5 daily 8-hr periods, leucine-1-<sup>14</sup>C and palmitate-9,10-<sup>3</sup>H were injected i.v. and the serum lipoprotein fraction was isolated. A significant decrease in the radioactivity of both lipid and protein moieties in the low density lipoproteins was noted. It was concluded that, as is the case in acute CCl<sub>4</sub> intoxication, chronic inhalation of CCl<sub>4</sub> leads to an accumulation of hepatic triglycerides and an inhibition of secretion of low density lipoproteins.

A GREAT deal of work has been done on the mechanism of production of fatty liver after carbon tetrachloride administration.<sup>1</sup> It has been demonstrated that the fatty liver resulting from acute intoxication with CCl<sub>4</sub> is secondary to a failure of the liver to secrete triglycerides in the form of low density lipoprotein.<sup>2-4</sup> The latter in turn may be due to an inhibition of protein synthesis<sup>5</sup> following a dislocation of the polyribosomes from the endoplasmic reticulum by CCl<sub>4</sub>,<sup>6</sup> either as a result of peroxidation of the lipid membranes<sup>7,8</sup> or a dissociation of the ribosomes due to a release of ribonuclease from lysozomes.<sup>9</sup>

Most of the data leading to theories of the action of CCl<sub>4</sub> have been obtained after acute administration of the toxin either i.p. into the gastrointestinal tract or with slices or subcellular particles. Relatively little work has been done on the effects of chronic CCl<sub>4</sub> intoxication, although industrial exposure to CCl<sub>4</sub> is usually chronic in nature. It has been established that chronic intoxication results in liver cirrhosis<sup>10,11</sup> with the alteration of various enzymes in serum. Thus serum prothrombin levels<sup>12</sup> and esterase activity<sup>13</sup> are decreased. In addition, increases in the rat serum levels of xanthine oxidase and serum glutamic oxaloacetic transaminase (SGOT)<sup>14</sup> and in rabbit serum glutamic pyruvate transaminase (SGPT), isocitric, glutamic, malic and lactic dehydrogenases, aldolase and phosphohexose isomerases were noted after chronic intoxication.<sup>15</sup>

<sup>\*</sup> Supported by grants from the Department of National Health and Welfare of Canada (604-7-306) and the Medical Research Council of Canada (MA-1266).

In contrast to the fatty liver and decreased lipoprotein secretion noted after acute administration of CCl<sub>4</sub>, chronic intoxication has been reported to cause a decrease in liver lipids<sup>16</sup> and an increase in serum lipids.<sup>17</sup> It is possible that the latter results are due to prolonged exposure times. In the light of the development of our understanding of the mechanism of fatty liver formation after acute CCl<sub>4</sub> intoxication, a study of the lipid levels and rates of secretion from the liver in rats exposed to CCl<sub>4</sub> daily over periods of 5 days was felt to be of value. The results of these investigations are presented in this communication.

### **METHODS**

Male hooded rats (obtained from Quebec Breeding Farms, St. Eustache, Quebec), weighing between 180 and 200 g and maintained on Purina chow, were used in all experiments. Animals were exposed in groups of four to mixtures of air and CCl<sub>4</sub> vapors for daily 8-hr periods in an exposure chamber consisting of a large desiccator equipped with a central air supply, an air exit and a water supply. The flow of air was measured by flowmeters placed before and after the exposure chamber and regulated with a needle valve. The CCl<sub>4</sub> was injected into the airstream at a steady rate by a dual infusion pump (Harvard Apparatus Inc., Dover, Mass.). Changes in the concentration of CCl<sub>4</sub> (ppm) were achieved by either increasing or decreasing the pump speed and the rate of air flow. The air supply line was constricted in order to increase the rate of air flow, thus insuring complete evaporation of the CCl<sub>4</sub>. Control animals were exposed under identical conditions except that no CCl<sub>4</sub> was added to the air supply line. It was found that the CCl<sub>4</sub>-exposed animals lost their appetite during exposure. Thus both the control and CCl<sub>4</sub>-exposed rats were deprived of food but allowed water *ad libitum* during the exposure periods.

During the study of lipid metabolism the animals were given, under nembutal anesthesia, an injection of 2·1 \(\mu\)mole albumin-bound palmitate-1-14C containing  $2 \times 10^6$  cpm/100 g body wt. via the jugular vein after the final 8-hr period of exposure. In some experiments, where it was desired to follow the secretion of the protein moiety of lipoproteins,  $1.2 \,\mu$ mole of L-leucine- $1^{-14}$ C containing  $2.6 \times 10^6 \,\mathrm{cpm}/100 \,\mathrm{g}$  body wt. was injected via the jugular vein, followed 30 min later by  $2.1 \mu$ mole albuminbound palmitate-9,10-3H containing  $2.6 \times 10^6$  cpm/100 g body wt. We have previously noted that under these conditions the maximum radioactivity levels in the lipid and protein moieties of the serum lipoproteins tend to coincide.<sup>18</sup> The first blood samples were then drawn 30 min after the palmitate injection. It has also been demonstrated that the lipoprotein secretion and turnover pattern in any individual animal is constant from week to week. 18 Therefore, all animals used in experiments in which the effect of chronic CCl<sub>4</sub> intoxication upon lipoprotein secretion was to be studied were first exposed for 5 days in air as controls and their lipoprotein secretion pattern was established. One week after the end of the control experiment, the same animals were subjected to a second exposure using CCl<sub>4</sub>.

Blood samples were collected from the tail vein, allowed to clot and the serum was removed quickly. Serum glutamic oxaloacetic transaminase activity was determined according to the method of Reitman and Frankel. Glycogen was isolated by precipitation with alcohol after digestion of the liver tissue in 30% KOH, then dissolved and reprecipitated several times and analyzed colorimetrically using the anthrone reagent. Liver lipids were extracted by the method of Folch et al. and free fatty

acids were removed by an alkaline wash. Neutral glycerides and phospholipids were separated on silicic acid columns and the ester bonds determined by the method of Rapport and Alonzo.<sup>22</sup> Serum triglycerides were isolated, counted and measured colorimetrically by the method of van Handel and Zilversmit<sup>23</sup> after removal of the free fatty acids by the Borgstrom technique.<sup>24</sup> Albumin-bound palmitate was prepared according to the procedure of Milstein and Driscoll.<sup>25</sup> Separation of serum lipoprotein fractions was carried out by the method of Havel et al.<sup>26</sup> in an International B-60 preparative ultracentrifuge with an SB-405 rotor and an average centrifugal force of 360,000 g. Aliquots of the lipoprotein fractions were dried on filter paper strips and the radioactivity was measured in a Tri-Carb liquid scintillation counter at a setting for the simultaneous counting of <sup>14</sup>C and <sup>3</sup>H with efficiencies of 64 and 10 per cent respectively.

Radioisotopes were obtained from New England Nuclear Corp., Boston, Mass. and SGOT kits from Sigma Chemical Company, St. Louis, Mo.

# RESULTS

In order to assess the degree of liver damage being caused by the chronic exposure to CCl<sub>4</sub>, the serum transaminase levels of animals exposed to 68 and 680 ppm CCl<sub>4</sub> vapor for daily periods of 8 hr were studied. The results shown in Fig. 1 indicate that,

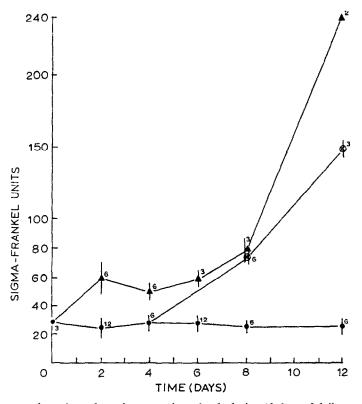
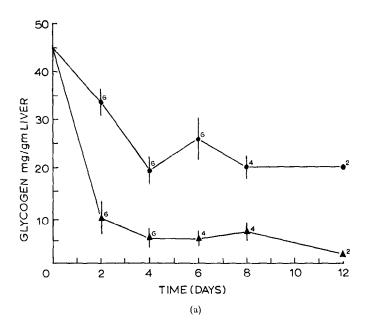


Fig. 1. Rat serum glutamic oxaloacetic transaminase levels during 12 days of daily exposure for 8 hr to CCl<sub>4</sub> vapors. The numbers on the curves refer to the number of animals; the vertical bars represent one S.E.M. 

■ Control; ⊗ = exposed to 68 ppm CCl<sub>4</sub>; ▲ = exposed to 680 ppm CCl<sub>4</sub>.

after exposure to 680 ppm CCl<sub>4</sub>, a rise in SGOT was noted after 2 days and that a further significant rise did not occur until the twelfth day of intoxication. With one-tenth the dose, no change in SGOT activity was noted for 4 days after which a rise in activity occurred.



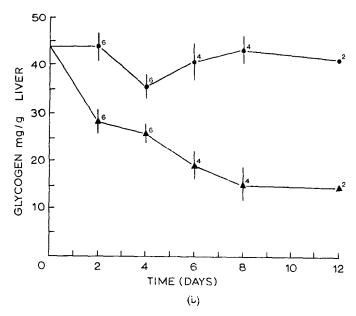
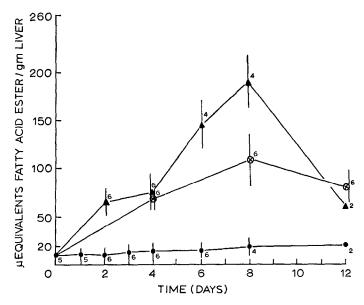


Fig. 2. Liver glycogen levels after daily exposure for 8 hr to CCl<sub>4</sub> vapors: (A) immediately after the last exposure; (B) 16 hr after the last exposure. The numbers on the curves refer to the number of animals; the vertical bars represent one S.E.M. ■ = Control; ▲ = exposed to 680 ppm CCl<sub>4</sub>.

We had earlier noted that one of the first changes following acute CCl<sub>4</sub> intoxication was a decrease in the level of liver glycogen.<sup>27</sup> The effect of the chronic intoxication upon the liver glycogen levels is shown in Fig. 2(a) and 2(b). Fig. 2(b) illustrates the fall in liver glycogen after successive days of exposure. It will be noted that the level of hepatic glycogen fell in both the intoxicated and control animals, stabilizing in the latter after 4 days. The glycogen level at the start of the experiment was obtained by fasting the animals for 8 hr in their cages. We assume that the stabilization of the glycogen at a lower, but constant, level represented an adaptation to the regular 8-hr periods of food deprivation. In the case of the animal exposed to 680 ppm CCl<sub>4</sub>, there was a marked fall in the level of glycogen with stabilization after 4 days at one-tenth the initial level. The curves of glycogen levels obtained with 68 ppm, not shown in the figure, were similar to that shown with 680 ppm. The influence of the 8-hr daily period of food deprivation compared to that of the CCl<sub>4</sub>-intoxication was further studied by measuring the glycogen levels of both control and exposed animals the following morning, after the animals had had 16 hr during the night for food consumption. The food intake of both the control and CCl4-exposed animals was similar during the nightly 16-hr periods. The results shown in Fig. 2(b) indicate that the control animals readily restored their glycogen levels to normal after each period of exposure. Under these conditions, the capacity of the intoxicated animals to deposit glycogen was diminished and the level was never restored to normal, although some glycogen was laid down.

Fatty livers are one of the prime results of acute CCl<sub>4</sub>-intoxication. Hence the level of liver lipids found after the chronic inhalation used in our experiments was studied. The results shown in Fig. 3 indicate that there is a rapid rise in the level of liver fatty acid esters in the liver, the level of lipid being related to the dose. However, with both



doses the level of lipid in the liver reached a maximum after 8 days; additional exposure resulted in a decrease. No significant change in the control liver lipid level was noted. Although not shown in the figure, the levels of lipid after the nightly 16-hr recovery period with food intake were somewhat higher than that seen immediately after exposure.

In order to study the handling of lipid by the livers from intoxicated animals, palmitate-1-14C was administered i.v. to rats after 5 days of daily 8-hr exposure to 68 ppm CCl<sub>4</sub>. The accumulation of the i.v. administered palmitate-1-14C in the neutral lipids of the liver at various time intervals is shown in Table 1. It will be noted that the radioactivity is greater in the livers from the CCl<sub>4</sub>-exposed animals at all the time intervals investigated. Between 30 and 60 min after the injection of

TABLE 1. INCORPORATION OF PALMITATE-1-14C INTO HEPATIC TRIGLYCERIDES\*

Time after palminate - injection (min)	Total incorporation			Sp. act.		
	Control (cpm/	CCl <sub>4</sub> -exposed mg liver $\pm$ S.E.N	P (1.)	Control (cpm/µg trigly	CCl <sub>4</sub> -exposed ceride ± S.E.M	P
30 60 180	$   \begin{array}{c}     163 \pm 5 & (8) \\     26 \pm 2 & (6) \\     32 \pm 6 & (4)   \end{array} $	240 ± 2 (10) 190 ± 38 (6) 101 ± 11 (4)	<0.001 <0.005 <0.01	$58.1 \pm 3.1  14.0 \pm 1.5  11.5 \pm 2.2$	$\begin{array}{c} 8.1 \pm 1.1 \\ 19.4 \pm 2.1 \\ 35.0 \pm 3.0 \end{array}$	<0.001 n.s. <0.001

<sup>\*</sup> Figures in parentheses represent the number of experiments; n.s. = not significant. Albumin-bound palmitate-1-14C injected i.v. immediately after the last of 5 periods of 8 hr of exposure to 68 ppm CCl<sub>4</sub> each.

palmitate, the isotope in the control animals had fallen by approximately 85 per cent while only a slight (25 per cent) decrease had occurred in the livers of the intoxicated animals. The lower sp. act. of the lipid in the livers of the CCl<sub>4</sub>-exposed animals compared to the controls 30 min after administration of the palmitate-1-14C was due to the large amount of lipid which had accumulated in the intoxicated livers during the 5 days of exposure to CCl<sub>4</sub>. After 60 min, sufficient labeled lipid had disappeared from the normal liver so that the sp. act. was not significantly different in the two groups of animals, and by 3 hr it was significantly higher in the CCl<sub>4</sub>-intoxicated animals. The incorporation into phospholipids was measured simultaneously but is not shown here, since there was no significant difference between the control and CCl4-exposed animals in either the incorporation or sp. act. of this fraction. The relationship between the accumulation and apparent retention of fatty acids as neutral lipids in the intoxicated liver and the secretion of radioactive triglycerides into the serum can be followed with the data in Fig. 4. It will be noted that there is a much lower rate of appearance of labeled triglycerides in the serum of the CCl<sub>4</sub>-intoxicated animals, the largest difference occurring within 30 min. However, due to the variation in rate of secretion among different animals it was not possible to obtain statistically significant differences. To overcome this, the experiment cited in Table 1 and Fig. 4 was repeated, each animal first undergoing 5 days of exposure to air alone as a control, then 1 week later being exposed to 680 ppm CCl<sub>4</sub>. At the end of each exposure period each animal received palmitate-9,10-3H and leucine-1-14C so that the rates of secretion of both the lipid and protein moieties of lipoproteins

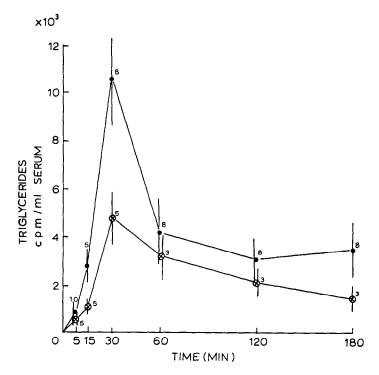


Fig. 4. Appearance of <sup>14</sup>C in serum triglycerides after injection of palmitate-1-<sup>14</sup>C in vivo immediately after 8 hr of exposure to CCl<sub>4</sub> vapors for 5 days. The numbers on the curves refer to the number of animals; the vertical bars represent one S.E.M. ■ = Control; ⊗ = exposed to 68 ppm CCl<sub>4</sub>.

could be followed simultaneously. It was found that a difference between the control and  $CCl_4$ -exposure periods occurred only in the low density lipoproteins (d < 1.064), although the high density lipoproteins and d > 1.21 proteins were also studied. The results are shown in Table 2 and indicate that there was a significant decrease in the radioactivity of the low density lipoproteins in the serum of the  $CCl_4$ -exposed

Table 2. Secretion of the lipid and protein moieties of the low density lipoproteins\*

Time after palminate injection (min)	Lipid incorporation			Protein incorporation		
	Control (cpm/ml seru	CCl <sub>4</sub> -exposed	P	Control	CCl <sub>4</sub> -exposed rum ± S.E.M.)	P
30 60 90 120	11,400 ± 1800 3200 ± 1400 1200 ± 300 520 ± 75	2400 ± 1300 1600 ± 800 1100 ± 300 450 ± 140	<0.005 n.s. n.s. n.s.	570 ± 47 360 ± 44 320 ± 54 260 ± 46	270 ± 54 211 ± 50 180 ± 19 180 ± 30	<0.005 <0.05 <0.05 n.s.

<sup>\*</sup> Each figure is the average of 5 experiments. Leucine-1- $^{14}$ C was injected i.v. immediately after the last of 5 daily 8-hr period of exposure to 680 ppm CCl<sub>4</sub> and albumin-bound palmitate-9,10- $^{3}$ H was injected 30 min later. n.s. = Not significant.

animals 30 min after the injection of the palmitate-9,10-3H. A decrease was also seen after 60 min, but this difference was no longer significant. The protein moiety of the serum lipoproteins of the poisoned animals was significantly decreased both at the 30- and 60-min interval. These data are therefore consistent with the view that a decrease in the secretion of low density lipoprotein has occurred.

### DISCUSSION

The results reported in this communication indicate that the hepatic changes following periodic carbon tetrachloride intoxication by inhalation over a 5-day period resemble those seen in acute CCl<sub>4</sub>-intoxication. Thus, as has previously been reported, after acute intoxication there is a decrease in the liver glycogen level. It is interesting that the loss in glycogen is not due solely to increased glycogenolysis but also involves an inability to resynthesize liver glycogen. The failure to restore the glycogen level to normal during the 16 hr after CCl<sub>4</sub> exposure is not due to a loss of appetite, since it was observed that food consumption was equal in the control and intoxicated rats. It is possible that glycogen synthetase is inhibited, as has previously been observed in liver slices exposed to CCl<sub>4</sub> vapors.<sup>28</sup> After chronic exposure to CCl<sub>4</sub>, the liver is characterized by an accumulation of neutral lipids which is not dose dependent until about 8 days. Subsequently there is a disappearance of lipid. The sharp rises in SGOT activity occur at this point. It is possible that these phenomena represent an alteration in the pathology from lipid deposition to necrosis. The separation of the lipid deposition and necrotic phases of CCl4 intoxication has previously been suggested after acute intoxication.29,30 Similar conclusions can be drawn from the experiments of Calvert and Brody,31 who demonstrated that antiadrenergic drugs inhibit necrosis but not accumulation of lipid after acute CCl<sub>4</sub> intoxication.

The data obtained with palmitate-1-14C suggest that, as is the case with acute intoxication, there is a failure of lipoprotein secretion. The accumulation of the labeled fatty acid esters in the liver is reflected by a decreased radioactivity in the serum triglycerides 30 and 60 min after exposure to CCl<sub>4</sub>. This corresponds to the significant diminution in the radioactivity of the lipid moiety of the lipoproteins after 30 min. Interpretation of the results is complicated by the observation that the specific activity of the triglycerides in the intoxicated liver is considerably reduced. Thus it is possible that the decrease in counts noted in the low density lipoproteins is simply due to a dilution of the isotope in the lipid moiety of the lipoprotein. However, this seems unlikely since there is also a significant decrease in the amount of leucine-14C incorporated into the protein moiety of the lipoproteins. These results lead to the same conclusion that has been suggested after acute intoxication with CCl<sub>4</sub>, namely that after chronic CCl<sub>4</sub> exposure there is a decrease in secretion of low density lipoproteins. The reason for this failure of lipoprotein secretion still remains to be determined.

## REFERENCES

- 1. R. O. RECKNAGEL, Pharmac. Rev. 19, 145 (1967).
- 2. R. O. RECKNAGEL, B. LOMBARDI and M. C. SCHOTZ, Proc. Soc. exp. Biol. Med. 104, 608 (1960).
- 3. H. M. Maling, A. Frank and M. G. Horning, Biochim. biophys. Acta 64, 540 (1962).
- 4. I. Weinstein, G. Dishmon and M. Heimberg, Biochem. Pharmac. 15, 851 (1966).
- 5. E. A. SMUCKLER and E. P. BENDITT, Biochemistry, N.Y. 4, 671 (1965).
- 6. E. A. SMUCKLER, O. A. ISERI and E. P. BENDITT, J. exp. Med. 116, 55 (1962).
- 7. T. F. SLATER, Nature, Lond. 209, 36 (1966).

- 8. R. O. RECKNAGEL and A. K. GHOSHEL, Lab. Invest. 15, 132 (1966).
- 9. D. H. Alpers and K. J. Isselbacher, Biochim. biophys. Acta 137, 33 (1967).
- 10. K. ATERMAN, Archs Path. 57, 1 (1954).
- 11. P. N. WAHI, H. D. TANDON and T. P. BHARADWAY, Archs Path. 62, 200 (1956).
- 12. J. L. BOLLMAN, Trans. Conf. Josiah Macy jr Fdn. Liver Injury 2, 18 (1944).
- 13. W. F. BALL and K. KAY, A.M.A. Archs ind. Hlth 14, 450 (1956).
- 14. W. D. BLOCK and H. H. CORNISH, Proc. Soc. exp. Biol. Med. 97, 178 (1958).
- 15. C. F. Fox, B. D. DINMAN and W. J. FRAJOLA, Proc. Soc. exp. Biol. Med. 111, 731 (1962).
- 16. R. E. STOWEL, C. S. LEE, K. K. TSUBOI and A. VILLASONA, Cancer Res. 11, 345 (1951).
- M. KOTANI, K. SEIKI, A. YAMASHITA, A. TAKASHIMA, T. NAKAGAWA and I. HORII, J. Lipid Res. 8, 181 (1967).
- 18. J. T. BUCKLEY, T. J. DELAHUNTY and D. RUBINSTEIN, Can. J. Biochem. 46, 341 (1968).
- 19. S. REITMAN and S. FRANKEL, Am. J. clin. Path. 28, 56 (1957).
- W. W. UMBREIT, R. H. BURRIS and J. H. STAUFFER, Manometric Techniques, pp. 239-78. Burgess, Minneapolis, Minn. (1959).
- 21. J. FOLCH, M. LEES and G. H. SLOANE-STANLEY, J. biol. Chem. 226, 497 (1957).
- 22. M. M. RAPPORT and N. ALONZO, J. biol. Chem. 217, 193 (1955).
- 23. E. VAN HANDEL and D. B. ZILVERSMIT, J. Lab. clin. Med. 50, 152 (1957).
- 24. B. Borgstrom, Acta physiol. scand. 25, 111 (1952).
- 25. S. W. MILSTEIN and L. H. DRISCOLL, J. biol. Chem. 234, 19 (1959).
- 26. R. J. HAVEL, H. A. EDER and J. H. BRAGDON, J. clin. Invest. 34, 1345 (1955).
- 27. A. J. MAXIMCHUK and D. RUBINSTEIN, Ann. occup. Hyg. 4, 49 (1961).
- 28. P. R. WELDON, B. RUBENSTEIN and D. RUBENSTEIN, Can. J. Biochem. 43, 647 (1965).
- 29. B. RUBENSTEIN and D. RUBINSTEIN, Can. J. Biochem. 42, 1268 (1964).
- 30. K. R. REES, K. P. SINHA and W. G. SPECTOR, J. Path. Bact. 81, 107 (1961).
- 31. D. N. CALVERT and T. M. BRODY, Am. J. Physiol. 198, 669 (1960).